

Molecular Methods: Choosing Appropriate Tests & Test Interpretation

David Rosenthal, DO, PhD
Attending Physician, Division of Allergy/Immunology
Northwell Health

Assistant Professor of Medicine and Pediatrics
Donald and Barbara Zucker School of Medicine at Hofstra/Northwell

January 23, 2018



Disclosures

- None



Learning Objectives

1. Understand the role of sensitivity/specificity in choosing the appropriate test
2. Understand variables which alter test results
3. Understand certain molecular methods used in the medical literature
4. Understand the importance of recoding data for test interpretation.



Question 1

An infant born to a woman infected with HIV has a positive Rapid HIV1 ELISA (3rd generation) at 2 months of age. What should you do first?

- A. Start HIV Antiretroviral medications as soon as possible.
- B. Repeat the Rapid HIV1 ELISA STAT.
- C. Order an HIV discriminatory test STAT.
- D. Repeat the HIV1 test test in 2 months.
- E. Recheck the specificity of the assay.

Question 2


- Your research coordinator tells you that the average fasting blood glucose in your healthy general pediatric patient population (n=253) is 125 mg/dl. What should you do for patients with high glucose?
 - A. Institute an evidence based medicine training session for all clinical staff to instruct them in proper diabetes screening.
 - B. Look at the data to understand why the patients all have high fasting glucose.
 - C. Begin treatment for DM
 - D. Order oral glucose tolerance tests
 - E. Order HgbA1c

Question 3

- Fifty percent of the QuantiFERON Gold Test-in-Tube (QFN) results are indeterminate. What should you do?
 - A. Repeat QFN on all patients with indeterminate results
 - B. Consider those patients with indeterminate results as screened for TB.
 - C. Call the lab to understand how the QFN test is done.
 - D. Refer all indeterminate patients to Peds ID for potential treatment.
 - E. Place a PPD on all of the patients with indeterminate results.

Selecting the right test: Statistics Reminders

- Sensitivity/Specificity
- Accuracy/Precision



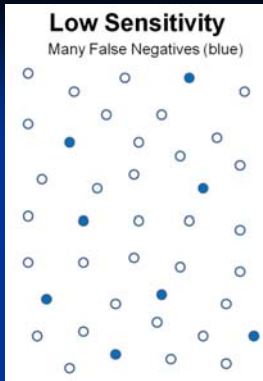
"Truth" or "Gold Std" test

	Pos	Neg	
Your test	Pos	a	b
	Neg	c	d
	a+c	b+d	Total a+b+c+d

- sensitivity = $a/(a+c)$
- specificity = $d/(b+d)$
- likelihood ratio (LR+) = sensitivity / (1-specificity) = $(a/(a+c)) / (b/(b+d))$
- likelihood ratio (LR-) = (1-sensitivity) / specificity = $(c/(a+c)) / (d/(b+d))$
- positive predictive value = $a/(a+b)$
- negative predictive value = $d/(c+d)$

Low Sensitivity

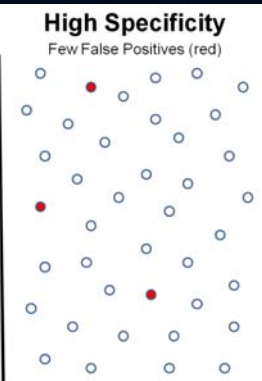
Many False Negatives (blue)



Failed Test

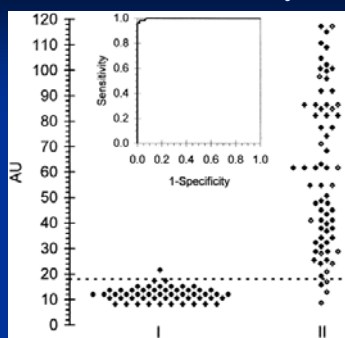
High Specificity

Few False Positives (red)



Passed Test

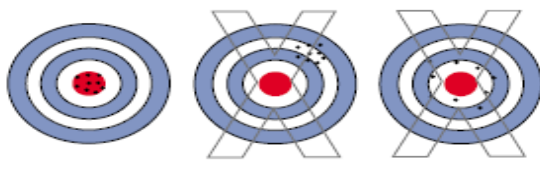
Laboratory Testing




Serum concentrations of IgA antibodies against TGc in the human TGc ELISA system in controls (I) and in patients having CD or DH (II). ◊, treated CD or DH patients; ♦, controls and untreated CD or DH patients. The chosen arbitrary cutoff for positivity (dashed line) is drawn at 18 AU. The ROC curve for the human TGc ELISA is shown as an inset.

Recombinant Human Tissue Transglutaminase ELISA for the Diagnosis of Gluten-sensitive Enteropathy. *Clinical Chemistry* December 1999 vol. 45 no. 12 2142-2149

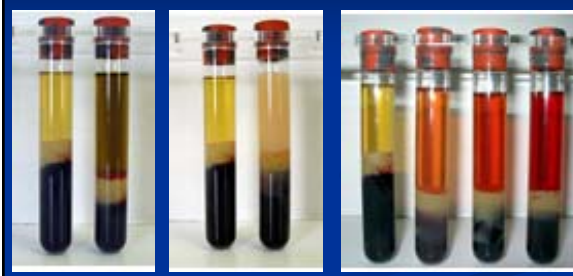
Accuracy and Precision




High accuracy
High precision High precision Low precision



Sample condition

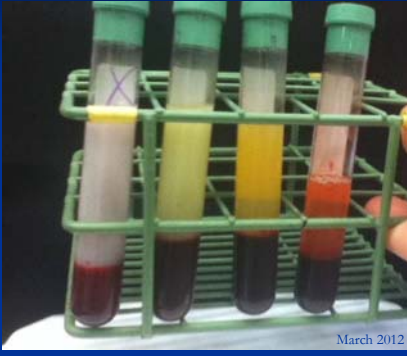


Volume of Tube
Transport temperature of Tube
Time from phlebotomy to testing




Sample Condition

Blood from a clinical trial patient. Obtained at the start of surgery.




March 2012



Selecting the right test

- Eg. Screening for Diabetes



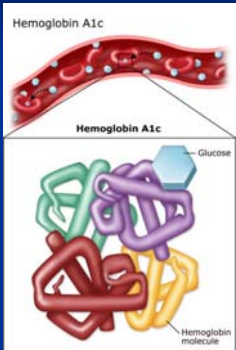

Diabetes Screening





- The right instrument



Diabetes Screening The Right Test





Oral glucose tolerance test

Selecting the right test


- Updated algorithms
- In vivo* vs. *In vitro*

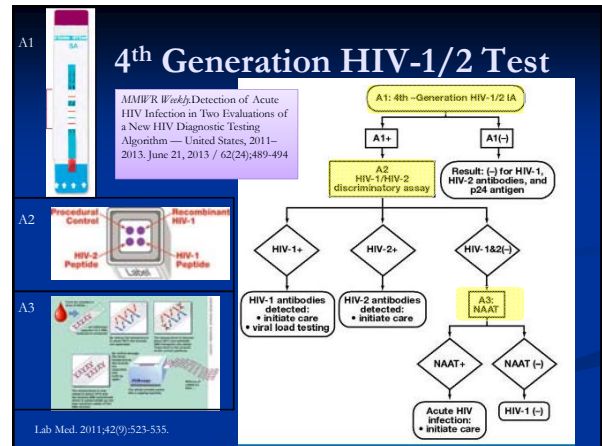
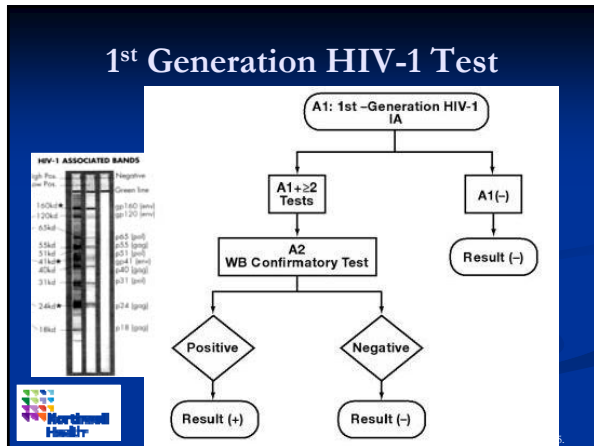


Development of HIV immunoassays

Generation	1st	2nd	3rd	4th
Antigen	[Diagram showing antigen binding to antibodies]			
Sample	[Diagram showing sample binding to antibodies]			
Conjugate	[Diagram showing conjugate binding to antibodies]			
Signal	[Diagram showing signal generation]			
Antigen	Lysate	Recombinant & synthetic		
Specificity	95-98%	>99%	>99.5%	99.5%
Sensitivity	99%	>99.5%	>99.5%	>99.8%
Window period	8-10 weeks	4-6 weeks	2-3 weeks	2 weeks
Immunoglobulin class detection	IgG	IgG	All	All
Approximate year of first release	1985	1987	1991	1997
Platforms	Plate assays Particle agglutination	Plate assays Automated generic platforms Particle agglutination Rapid assays	Plate assays Dedicated instruments Rapid assays	Plate assays Dedicated instruments Rapid assays in development

Future Microbiol. 2009;4(8):963-982.
http://www.medscape.com/viewarticle/715166_2

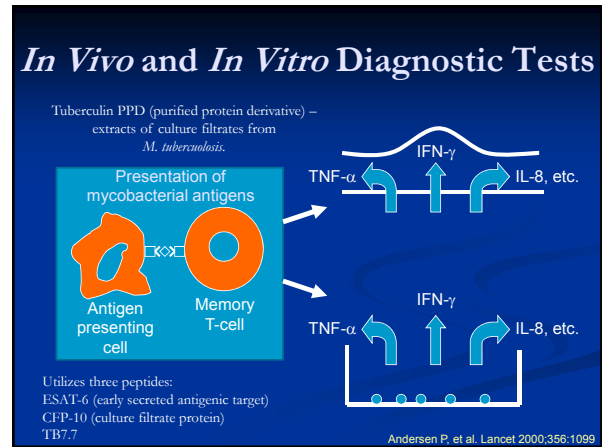




Case: Newborn

Ex-36 wk, SGA (BW 1906 kg) born to a G6P21112 mother via C/S due to late decels

Test	Mother	Baby
1 st TM HIV EIA	Neg (Oct, Nov 2013)	
3 rd TM HIV EIA	Neg (5/20/14)	
L&D HIV Rapid EIA	Pos x 2 (5/23/14)	
4 th Generation HIV	Neg (5/23/14)	
HIV Viral Load	Pos: 118,000 RNA copies	
HIV Multispot	Neg	
HIV 4 th Generation EIA		Neg
HIV Newborn screen		Neg
HIV Newborn NYS labs		Neg Multispot, Neg PCR
HIV Viral Load	Neg (5/27/14, 5/29/14)	Neg (5/26/14, 5/27/14)



Tuberculin Skin Testing Mantoux Method

5 TU of PPD → 48 to 72 hours

Interpretation depends on Patient Risk Factors

Induration ≥5 mm	Induration ≥10 mm	Induration ≥15 mm
<ul style="list-style-type: none"> HIV-infected Recent contact with a person with TB disease Fibrotic changes on CXR c/w with prior TB H/O organ transplants Immunosuppressed for other reasons (e.g., taking the equivalent of >15 mg/day of prednisone for 1 month or longer, taking TNF-α antagonists) 	<ul style="list-style-type: none"> Recent immigrants (< 5 yrs) from high-prevalence country Injection drug users Residents and employees of high-risk congregate settings Mycobacteriology laboratory personnel Clinical conditions that place them at high risk Children < 4 years of age Infants, children, and adolescents exposed to adults in high-risk categories 	<ul style="list-style-type: none"> any person including persons with no known risk factors for TB.

Whole Blood IFN-γ Assay QuantiFERON-TB Test

Stage 1 Whole Blood Culture


Draw blood into special tubes → Incubate → IFN-γ is released from sensitized T-cells

Stage 2 IFN-gamma ELISA

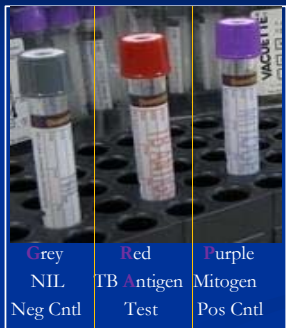
Centrifuge to separate plasma from cells → Measure [IFN-γ] in 'Sandwich' ELISA → Computerized interpretation

QuantIFERON®-TB Gold Test Advantages and Disadvantages


- Advantages:
 - Only one visit required
 - Objective and reproducible; not operator-dependent
 - No cross reactivity with BCG, little cross-reactivity with non-tuberculous mycobacteria
 - Controls for low or no immune response
 - No chance of ulceration due to brisk skin test reaction
- Disadvantages:
 - Blood must be received in lab within 16 hours
 - Labor intensive for the lab
 - Not much data for some patient groups



QuantIFERON –TB Gold In-Tube





- All tubes have a white ring on the top of the cap.
- Draw samples in this order! “CRAB.”
- Collect 1 mL of blood directly into each of the Quantiferon-TB Gold IT blood collection tubes.
- Should always be drawn last if other blood samples are being collected.
- Blood draws slowly into tubes. Keep the tube on the needle for 2-3 seconds once the tube appears to be filled.




Interpreting the Results

- Amount of IFN- γ (IU/mL) is reported from 3 tubes:
 - Nil
 - TB Antigen
 - Mitogen
- Calculations:
 - TB Antigen-Nil
 - Mitogen-Nil
- Analysis
 - Positive
 - Negative
 - Indeterminate


Interpretation of QTF-TB Gold In-Tube Test

- Tubes not shaken enough – PHA not solubilized
- Increased time (>16 hrs) from blood draw to incubation (36-38°C) in lab
- Storage of filled blood tubes outside of temp range (22°C±5°C)

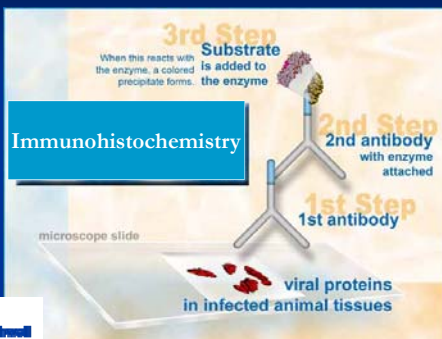



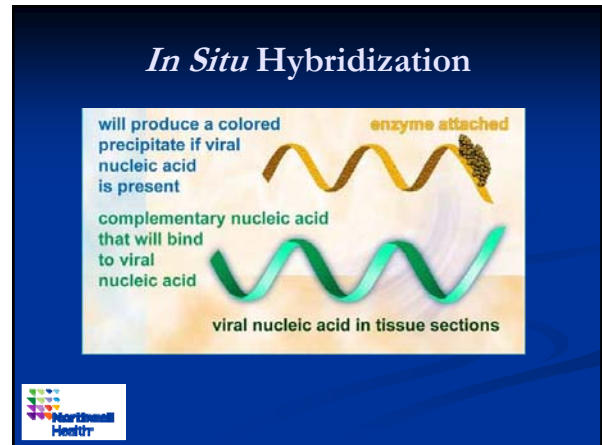
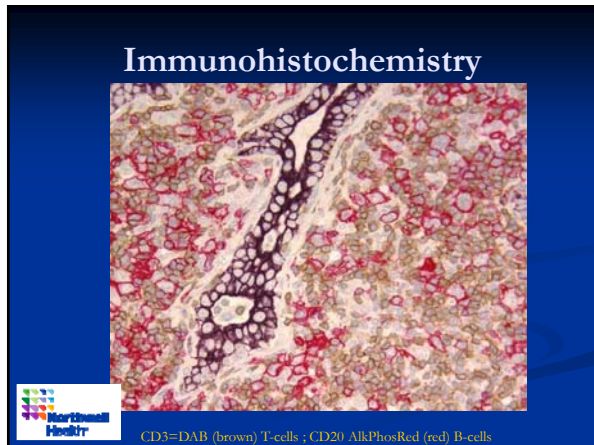
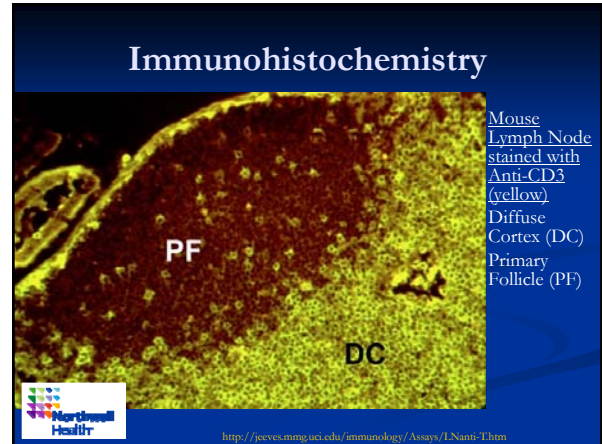
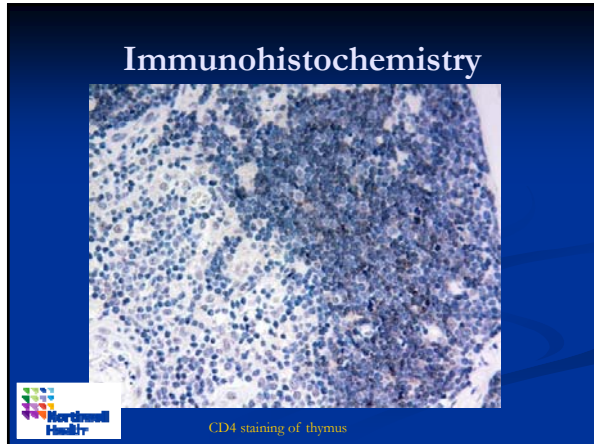
Selecting the right Molecular Method

- Immunohistochemistry vs In situ hybridization
- PCR vs. Quantitative PCR
- Understanding ELISA results - “titers”
- Blotting – Western, Northern, Southern...



Immunohistochemistry



Immunohistochemistry vs. In Situ Hybridization

- Both help you find which cells are making your gene/protein of interest
- Immunohistochemistry uses an antibody to find a protein in the cell
- In Situ Hybridization classically uses DNA or RNA (oligonucleotides) to find nucleic acids in the cell.

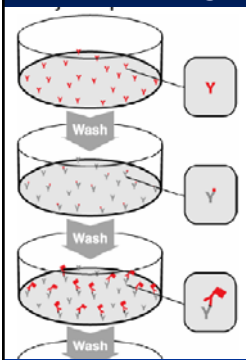
ELISA



ELISA measures the amount of antigen in liquid phase
 ELISPOT measures the number of cells with antigen on them




ELISA – Ag in fluids



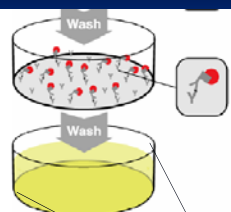
The assay is commonly performed in 96-well microtiter plates. In the first step the wells are coated with a monoclonal antibody (Y) specific for the cytokine to be determined.

Samples containing unknown amounts of cytokine (•) are added to the wells. To other wells, cytokine standard of known concentration is added. During the incubation, the cytokine is captured by the antibody attached to the wells.

The sample/standard is removed by washing and a biotinylated antibody (X) is added.

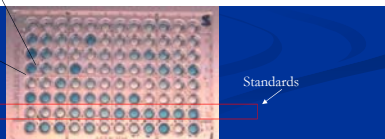


ELISA - Ag in fluids




Streptavidin-enzyme (ALP or HRP) conjugate (A) is added.

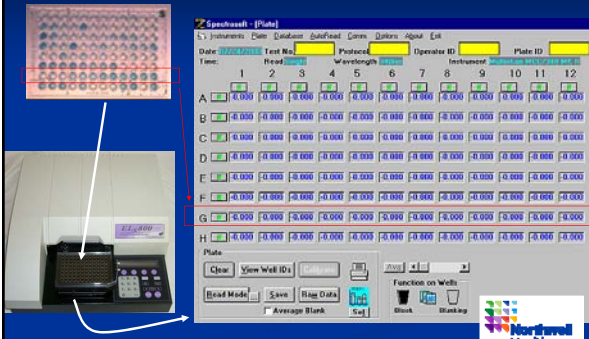
Finally, a chromogenic substrate for ALP or HRP is added and the plates are developed until colour emerges. Within the detection stage of the assay, the intensity of the colour is directly proportional to the amount of cytokine added to each well. The concentration of samples is determined by comparison to the cytokine standard.




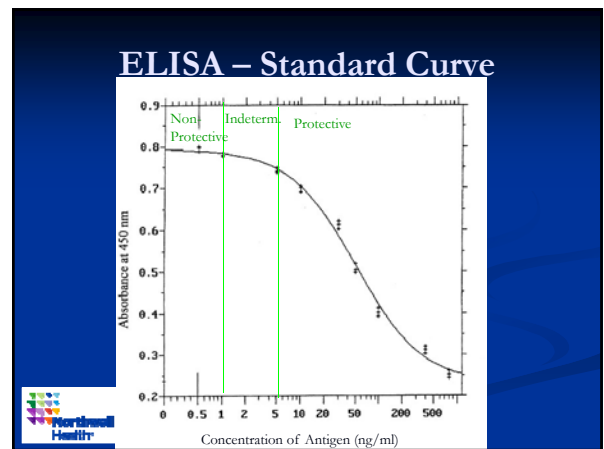
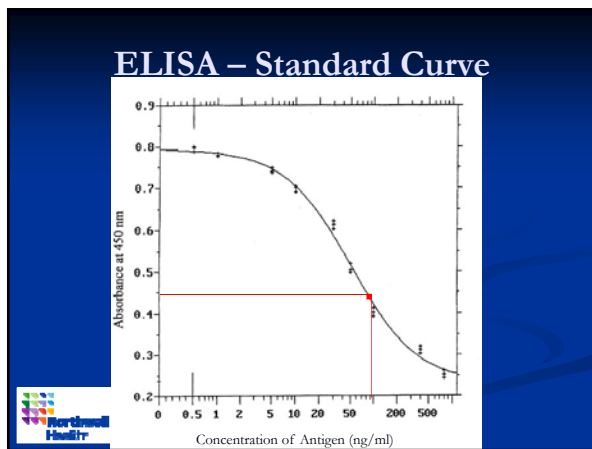
Standards

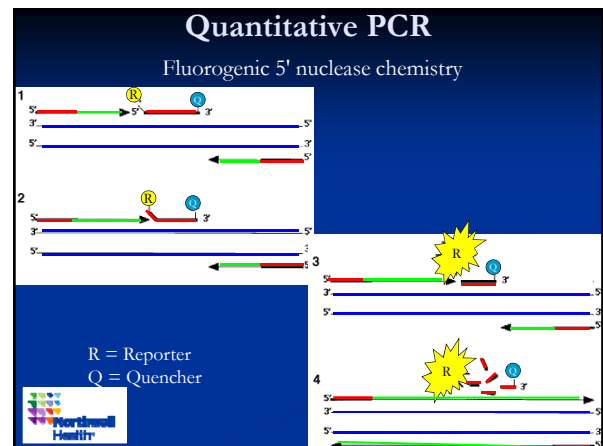
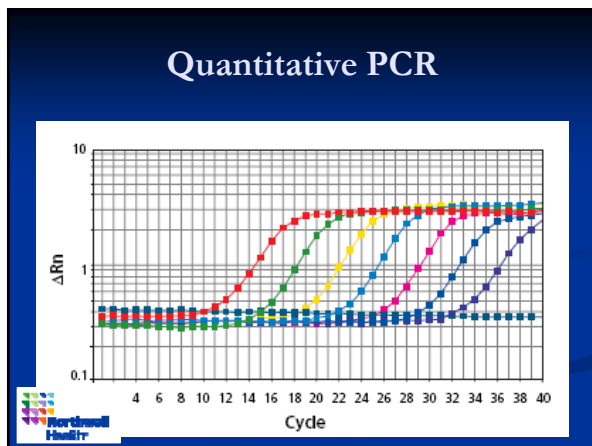
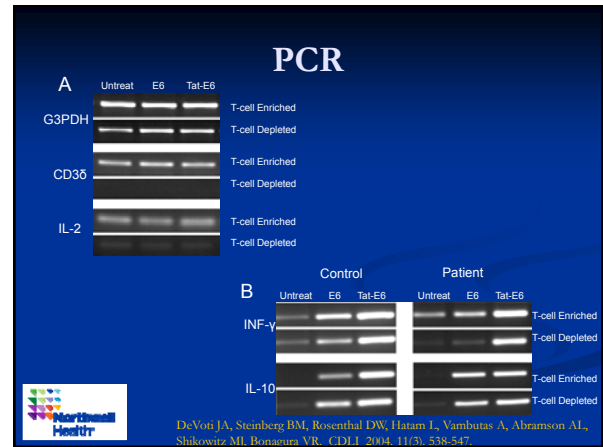
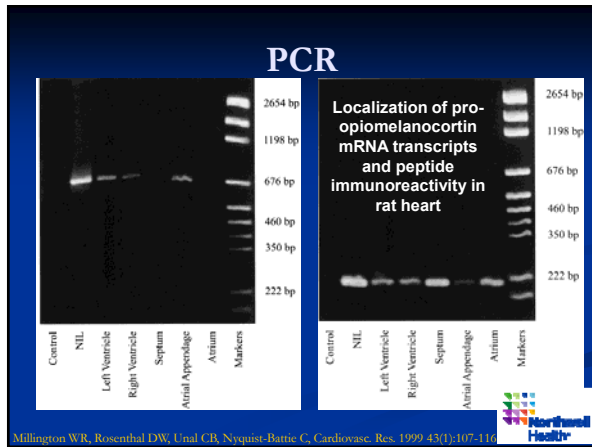
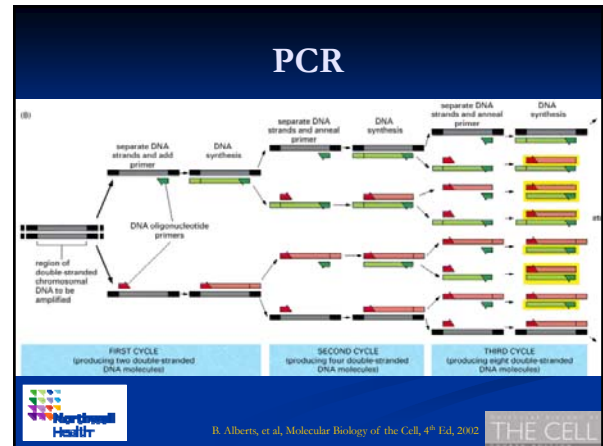
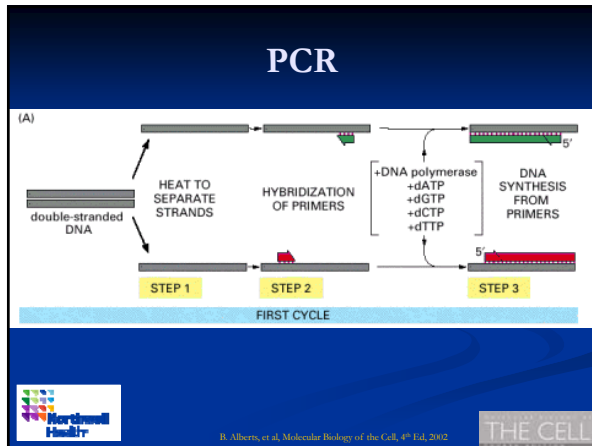


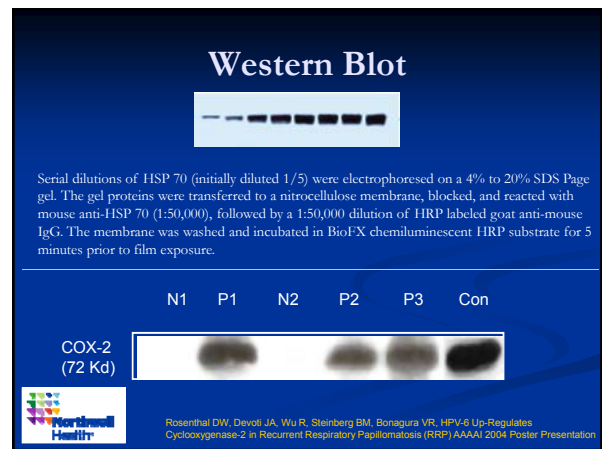
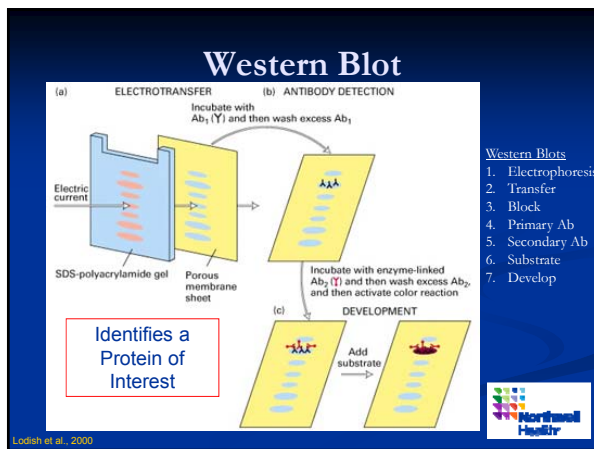
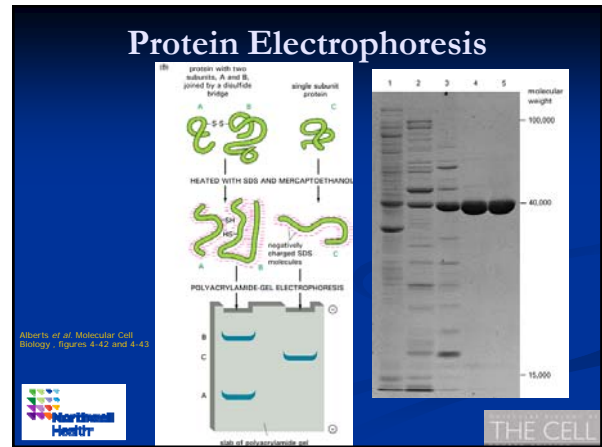
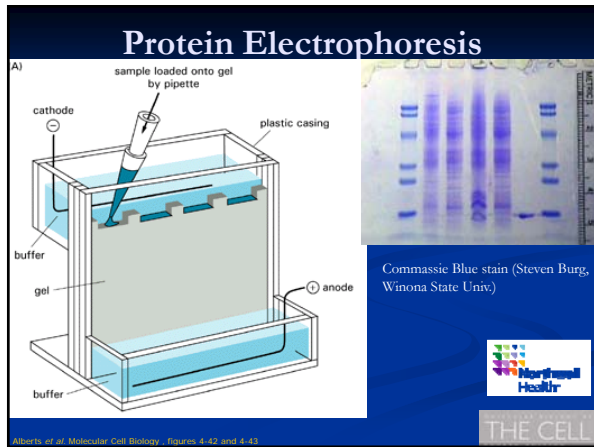
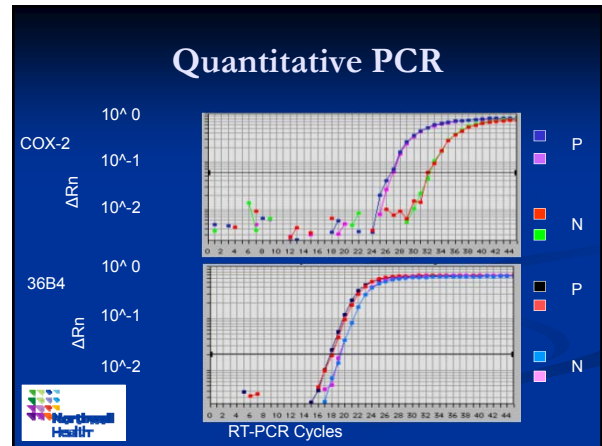
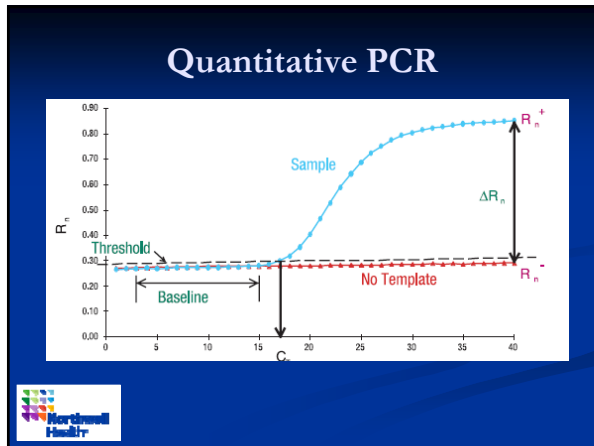
ELISA

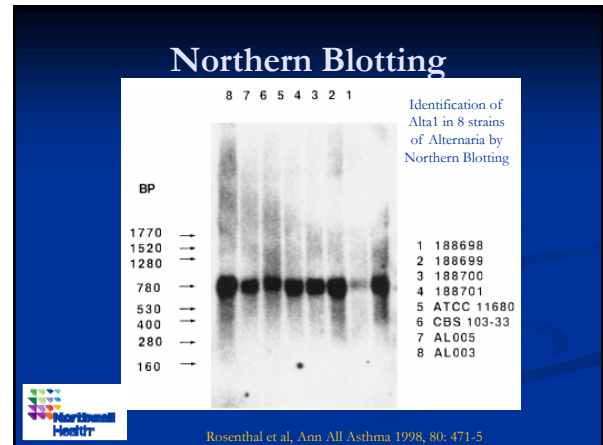
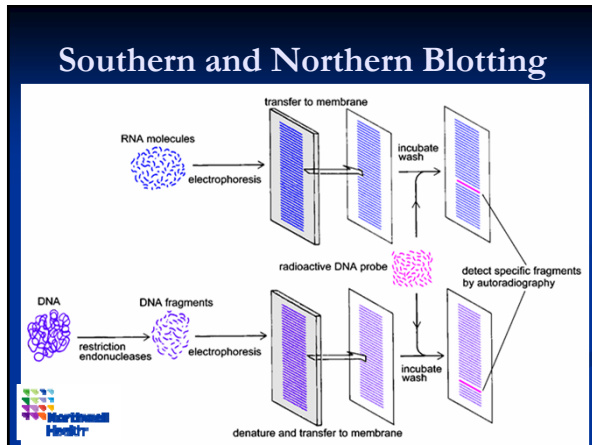
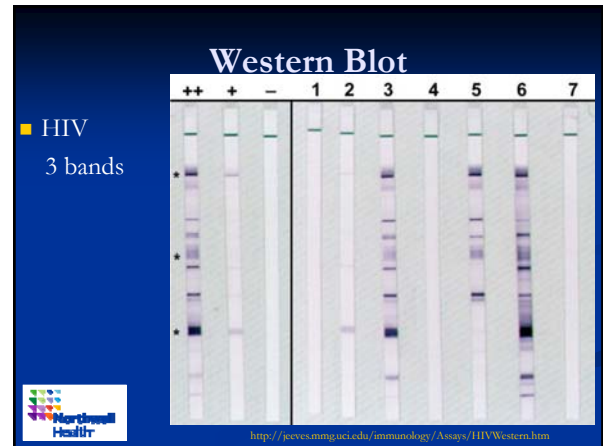
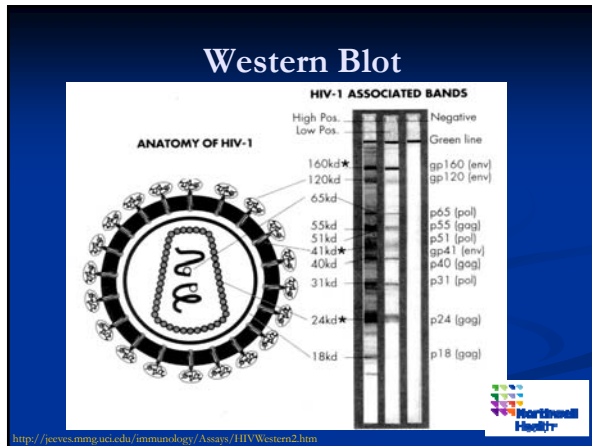


Time	1	2	3	4	5	6	7	8	9	10	11	12
A	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
B	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
C	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
D	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
E	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
F	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
G	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
H	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000







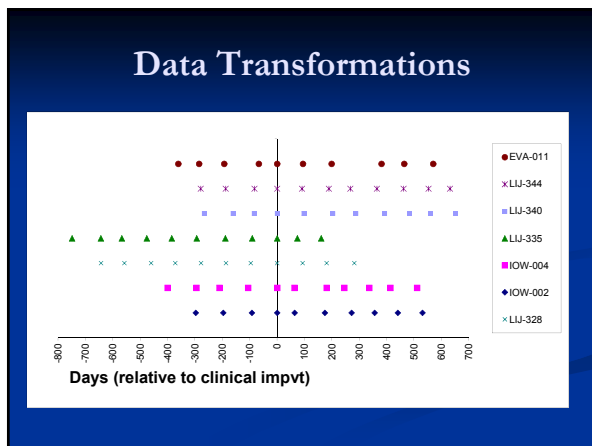
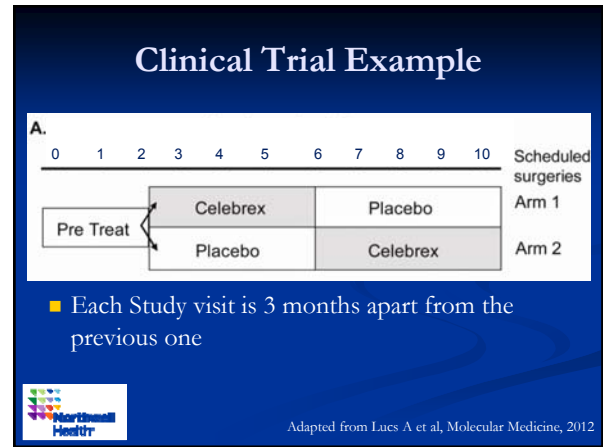
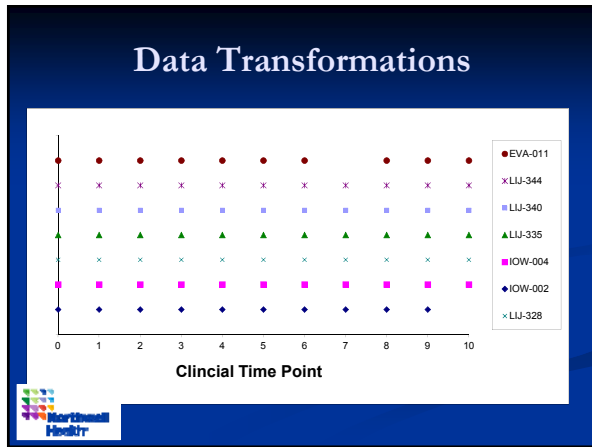
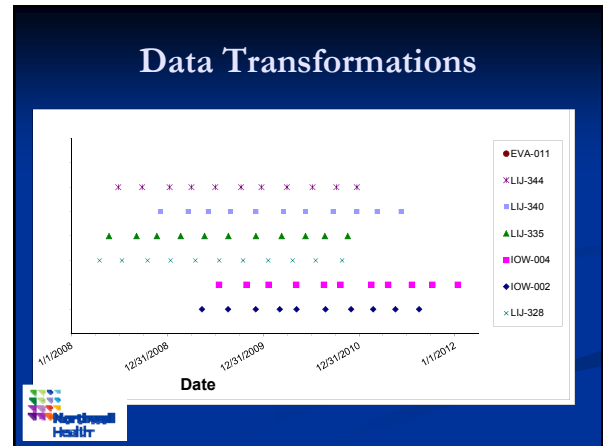
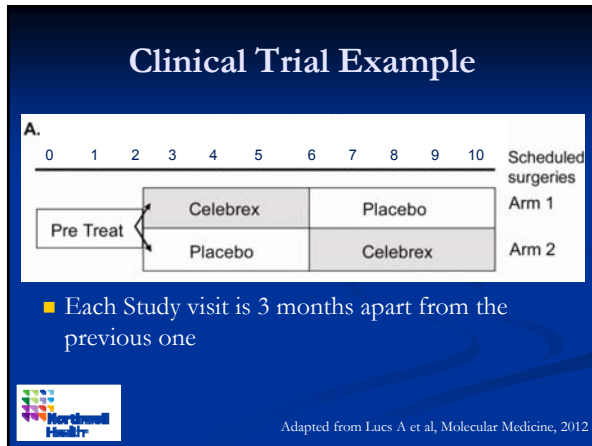
Blotting... how to avoid confusion

- Dr. EM **Southern** first described “Detection of specific sequences among DNA fragments separated by gel electrophoresis.” (J Mol Biol. 1975)
- Thus it was Blotting is described as:
 - Southern=DNA
 - Northern=RNA
 - Western=Protein

<http://peves.mmg.uci.edu/immunology/Assays/SouthernNorthern.htm>

Study Design and Interpreting the Data

<http://peves.mmg.uci.edu/immunology/Assays/SouthernNorthern.htm>



Question 1

An infant born to a women infected with HIV has a positive Rapid HIV1 ELISA (3rd generation) at 2 months of age. What should you do first?

- A. Start HIV Antiretroviral medications as soon as possible.
- B. Repeat the Rapid HIV1 ELISA STAT.
- C. Order an HIV discriminatory test STAT.
- D. Repeat the HIV1 test test in 2 months.
- E. Recheck the specificity of the assay.

Question 2

Your research coordinator tells you that the average fasting blood glucose in your healthy general pediatric patient population (n=253) is 125 mg/dl. What should you do for patients with high glucose?

- A. Institute an evidence based medicine training session for all clinical staff to instruct them in proper diabetes screening.
- B. Look at the data to understand why the patients all have high fasting glucose.
- C. Begin treatment for DM
- D. Order oral glucose tolerance tests
- E. Order HgbA1c

Question 3

Fifty percent of the Quantiferon Gold Test-in-Tube (QFN) results are indeterminate. What should you do?

- A. Repeat QFN on all patients with indeterminate results
- B. Consider those patients with indeterminate results as screened for TB.
- C. Call the lab to understand how the QFN test is done.
- D. Refer all indeterminate patients to Peds ID for potential treatment.
- E. Place a PPD on all of the patients with indeterminate results.

Northwell Health Division of
Allergy/Immunology

Adult, Adolescent and
Pediatric
Allergy, Asthma, and
Clinical Immunology

- HIV/AIDS (Age ≤ 30 years old)
- Primary Immune Deficiency
- Allergic Rhinitis
- Urticaria / Angioedema
- Food / Medication Allergy
- Eczema/Atopic Dermatitis
- Asthma



David Rosenthal, D.O. Ph.D.
Northwell Health
Division of Allergy/Immunology
865 Northern Blvd, Suite 101
Great Neck, NY 11021
✉: drosenthal@northwell.edu

☎: 516-622-5070
☎: 516-622-5060